

Name and Address of Exporter AUSTRALIA		Name and Address of Importer NEW ZEALAND	
		Import Permit N°	
Description of Animal Reproductive Material			
<u>Number</u>	<u>Kind (Species and type; eg bovine semen)</u>	<u>Condition (Fresh/Frozen)</u>	<u>Identification (straw numbers, packing list)</u>
	OVINE/CAPRINE SEMEN	FROZEN STRAWS	SEE ATTACHED

I, Dr, an approved Centre Veterinarian of (Name of semen collection centre), declare that the goods described in the following pages have complied with the importing country requirements.

Signature (pdf. doc only)

Date

ZOOSANITARY CERTIFICATE

Commodity: OVINE/CAPRINE SEMEN
To: NEW ZEALAND

Import Permit Number:

Exporting Country: AUSTRALIA

Competent Authority: DEPARTMENT OF AGRICULTURE, WATER AND THE ENVIRONMENT ("THE DEPARTMENT")

I. INFORMATION CONCERNING THE DONOR ANIMALS AND SEMEN

See attached Schedule

Total number of straws/vials:

II. ORIGIN OF THE SEMEN

Name, address and approval/registration number of semen collection centre:

Name and address of owner:

Name and address of exporter:

III. DESTINATION OF SEMEN

Name and address of importer:

SANITARY INFORMATION**1 Donor animals and semen collection centre**

1.1 The donor animals:

Either: 1.1.1 were born in and lived continuously in Australia

Or: 1.1.2 were imported into Australia from New Zealand.
(Delete as appropriate)

1.2 For any semen for export to New Zealand of a bloodline derived from semen or embryos imported from any country other than New Zealand or South Africa, the semen originated from animals that were conceived (using non-imported semen), born and lived continuously in Australia. The donor animal's parents were also conceived (using non-imported semen), born and lived continuously in Australia.

1.3 The flock(s) of origin of the donor animals and the semen collection centre were free from any quarantine restrictions from 90 days before the first semen collection until completion of the testing of the donor animals as required by this certificate.

1.4 The donor animals were held in the semen collection centre for a continuous period of at least 30 days before the collection of semen for this consignment and until the testing specified in this certificate was completed. During this time they were not used for natural mating and were isolated from animals not of equivalent health status.

1.5 The centre is approved by the Department for collection of semen for export, and inspected at least annually during the breeding season by an Official Veterinarian.
Date of last inspection: **See attached Schedule**

2. Semen collection

2.1 The period of semen collection(s) for this consignment was 60 days or less.

2.2 On the day(s) of collection of semen, the donor animals were examined by the team veterinarian and were free from any clinical evidence of infectious diseases caused by micro-organisms transmissible in semen.

2.3 The semen was collected, processed and stored under the supervision of a Department approved semen collection centre veterinarian in accordance with the OIE *Code*, Appendix for small ruminant semen.

2.4 Antibiotics effective against *Leptospira* and *Mycoplasma* spp. were added to the diluent. The names and concentrations of antibiotics included in the semen diluent are as follows:
See attached Schedule

3. Testing and treatment of donor animals

(NB: indicate which option was followed, test used and date(s) of sampling)

3.1 For bluetongue virus (BT):

Either 3.1.1 When importing from BT virus free zones (as defined by the OIE *Code*):

Either 3.1.1.1 The Donor animals were kept in a BT free zone for at least the 100 days prior to, and during, collection of the semen;

Or 3.1.1.2 The donor animals were subjected to serological tests to detect antibodies to BT, such as the competitive ELISA or the agar gel immunodiffusion test (AGID), between 28 and 60 days after the last collection for this consignment, with negative results;

Or 3.1.1.3 The donor animals were subjected to tests for BT, such as a virus isolation test or a polymerase chain reaction (PCR) test on blood samples collected at commencement and conclusion of, and at least every 7 days (for virus isolation

test) or at least every 28 days (for PCR test) during, semen collection for this consignment, with negative results.

Test used:

Date(s) of sample collection:

Or 3.1.2 When importing from BT virus seasonally free zones (as defined by the OIE Code):

Either 3.1.2.1 The donor animals were kept during the seasonally free period in a BT virus seasonally free zone for at least the 100 days prior to commencement of, and during, semen collection;

Or 3.1.2.2 The donor animals were subjected to serological tests to detect antibodies to BT, such as the competitive ELISA or the AGID test, between 28 and 60 days after the final collection for this consignment, with negative results;

Or 3.1.2.3 The donor animals were subjected to tests for BT, such as a virus isolation test or a PCR test on blood samples collected at commencement and conclusion of, and at least every 7 days (for virus isolation test) or at least every 28 days (for PCR test) during, semen collection for this consignment, with negative results.

Test used:

Date(s) of sample collection: .

Or 3.1.3 When importing from BT virus infected zones (as defined by the OIE Code):

Either 3.1.3.1 The donor animals were protected from *Culicoides* attack for at least the 100 days prior to commencement of, and during, semen collection;

Or 3.1.3.2 The donor animals were subjected to serological tests to detect antibodies to BT, such as the competitive ELISA or AGID test, between 28 days and 60 days after the final collection for this consignment, with negative results;

Or 3.1.3.3 The donor animals were subjected to tests for BT, such as a virus isolation test or PCR test on blood samples collected at commencement and conclusion of, and at least every 7 days (for virus isolation test) or at least every 28 days (for PCR test) during, semen collection for this consignment, with negative results.

Test used:

Date(s) of sample collection:

(Delete as appropriate)

3.2 For Q fever: Between 10 and 30 days after the final collection of semen for export to New Zealand, the donor animals were tested with negative results for Q fever using either the complement fixation test (CFT) (negative being no fixation of complement at a dilution of 1:10 or higher) or the ELISA.

Test used:

Date of sample collection:

3.3 All testing was conducted at a laboratory approved by the Department to conduct export testing and laboratory results for tests specified in this certificate are attached.

4 Storage and transport

4.1 All straws are clearly marked with the identification of the donor animal and the date of collection. If a code is used for this information, its decipher must accompany the consignment.

4.2 The semen was stored only with other semen or embryos that were eligible for export to New Zealand. The containers were held in an approved storage place under the supervision of the Department until export.

4.3 The semen was placed in new or sterilised transport containers filled with fresh (previously unused) liquid nitrogen.

Method of sterilisation (if applicable):

Date of sterilisation (if applicable):

~~4.4 Prior to export, the container in which the semen is to be transported was sealed by either the semen collection centre veterinarian or an Official Veterinarian, using seals bearing the marks:~~

Schedule of Donors

Donor Animal/Semen					
Breed	Identification	D.O.B	Collection Date(s)	Straw Identification	Number of Straws
Antibiotics Added to Diluent:					
Bluetongue Testing			Q Fever Testing		
BT Option	Test Type	Date	Test Type	Date	Result
Semen Collection Centre					
Name		Address		Registration No:	Last Inspection
Owner					
Name		Address			

Signature (pdf. doc only)

Date