

Name and Address of Exporter		Name and Address of Importer	
AUSTRALIA		NEW ZEALAND	
		Import Permit N^o	
Description of Animal Reproductive Material			
<u>Number</u>	<u>Kind (Species and type; eg bovine semen)</u>	<u>Condition (Fresh/Frozen)</u>	<u>Identification (straw numbers, packing list)</u>
	OVINE / CAPRINE EMBRYOS	FROZEN STRAWS	SEE ATTACHED

I, Dr, an approved Embryo Transfer Veterinarian, declare that the goods described in the following pages have complied with the importing country requirements.

.....
Signature (pdf. doc only)

.....
Date

Commodity: OVINE / CAPRINE EMBRYOS

To: NEW ZEALAND

Import Permit Number:

Exporting Country: AUSTRALIA

Competent Authority: DEPARTMENT OF AGRICULTURE AND WATER RESOURCES ("the Department")

I. INFORMATION CONCERNING THE DONOR ANIMALS (FEMALES AND MALES)

(For more than one animal, please use a schedule)

	Breed	Identification	Date of Birth	Datch number of semen (if applicable) or date of natural mating
Donor ewe/ doe				
Donor ram/buck				

Name, address and approval/registration number of embryo collection centre(s):

Name and address and approval/registration number of semen collection centre (if applicable):

Name and address of owner:

II. INFORMATION CONCERNING THE OVINE/CAPRINE EMBRYOS

	Date(s) of collection	Straw Identification (markings to be indelible)	No. of straws per donor (include the no. of embryos per straw)
Embryos	<i>Refer to Attach schedule</i>		

Total number of embryos in this consignment:

Name and address of exporter:

III. DESTINATION OF THE EMBRYOS

Name and address of importer:

IV. SANITARY INFORMATION

1. Donor animals and embryo collection centre

1.1 The donor females (delete as appropriate):

Either: 1.1.1 were born in and lived continuously in Australia;

Or: 1.1.2 were imported into Australia from New Zealand.

1.2 Embryos for export to New Zealand of a bloodline derived from semen or embryos imported from any country other than New Zealand or South Africa, originated from animals that were conceived (using non-imported semen), born and lived continuously in Australia.

1.3 The flock(s) of origin of the donor females and the embryo collection centre, were free from any quarantine restrictions from 60 days before the first embryo collection until completion of the donor animal testing for this consignment.

1.4 The donor females were held in a Department approved embryo collection centre for a continuous period of at least 30 days before the collection of embryos for this consignment and until the testing specified in this certificate was completed. During this time were isolated from animals not of an equivalent health status.

1.5 The ova were fertilised as follows (delete as appropriate):

Either 1.5.1 Donor males used for natural service were of an equivalent isolation and tested health status to the donor females

Or 1.5.2 Semen used for insemination was eligible for export to New Zealand.

1.6 The embryo collection centre is approved by the Department to collect embryos for export and is inspected by an Official Veterinarian at least once a year, at a time when embryo collection is being conducted.

Date of last inspection:

2. Embryo collection and processing

2.1 The period of embryo collection(s) for this consignment was 60 days or less.

2.2 On the day(s) of collection of the embryos, all female donors were examined by the team veterinarian and were free from any clinical evidence of infectious diseases caused by micro-organisms transmissible in embryos.

2.3 The embryos were collected, processed and stored under the supervision of a Department approved embryo collection team veterinarian in accordance with the OIE *Code*, Appendix for *in vivo* derived embryos.

2.4 The embryos were collected, washed, processed, identified and stored under conditions that comply with the recommendations in the *Manual of the International Embryo Transfer Society*. Each embryo had an intact zona pellucida and was examined over its entire surface at not less than 50X magnification and was free of adherent material.

2.5 The embryos were treated with the enzyme trypsin in accordance with the recommendations of the IETS *Manual*.

2.6 All biological products of animal origin used in the media and solutions for collection, processing, washing or storage of embryos were free of pathogenic organisms including pestiviruses. Media and solutions were sterilised according to the IETS *Manual* and handled in such a manner as to ensure that sterility was maintained. Antibiotics effective against *Leptospira spp.* were added to collection, processing, washing

and storage media as recommended in the IETS *Manual*, or a combination of antibiotics with equivalent activity was used.

2.7 The names and concentrations of antibiotics used in the embryo preparations included:

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2.8 Only frozen *in-vivo* fertilised ovine / caprine embryos are included in this consignment.

3 Testing and treatment of donor animals

3.1 For bluetongue virus (BT) virus:

(NB: indicate which option was followed, test used and date(s) of sampling)

Either 3.1.1 When importing from BT virus free zones (as defined by the OIE Code):

Either 3.1.1.1 The donor animals were kept in a BT free zone for at least 100 days prior to, and during, collection of the embryos;

Or 3.1.1.2 The donor animals were subjected to serological tests to detect antibodies to BT, such as the competitive ELISA or the agar gel immunodiffusion test (AGID), between 28 and 60 days after the final collection for this consignment, with negative results;

Or 3.1.1.3 The donor animals were subjected to tests for BT, such as a virus isolation test or a polymerase chain reaction (PCR) test, on blood samples taken on the day(s) of embryo collection for this consignment, with negative results.

Test used:

Date(s) of sample collection:

Or 3.1.2 When importing from BT virus seasonally free zones (as defined by the OIE Code):

Either 3.1.2.1 The donor animals were kept during the seasonally free period in a BT virus seasonally free zone for at least 100 days prior to commencement of, and during, embryo collection;

Or 3.1.2.2 The donor animals were subjected to serological tests to detect antibodies to BT, such as the competitive ELISA or the agar gel immunodiffusion test (AGID) test, between 28 and 60 days after the final collection for this consignment, with negative results;

Or 3.1.2.3 The donor animals were subjected to tests for BT, such as a virus isolation test or a polymerase chain reaction (PCR) test, on blood samples taken on the day(s) of embryo collection for this consignment, with negative results.

Test used:

Date(s) of sample collection:

Either 3.1.3 When importing from BT virus infected zones (as defined by the OIE Code):

Either 3.1.3.1 The donor animals were protected from *Culicoides* attack for at least 100 days prior to commencement of, and during, embryo collection;

Or 3.1.3.2 The donor animals were subjected to serological tests to detect antibodies to BT, such as the competitive ELISA or the agar gel immunodiffusion test (AGID); between 28 and 60 days after the final collection for this consignment, with negative results;

Or 3.1.3.3 The donor animals were subjected to tests for BT, such as a virus isolation test or a polymerase chain reaction (PCR) test, on blood samples taken on the day(s) of embryo collection for this consignment, with negative results.

Test used:

Date(s) of sample collection:

(Delete as appropriate)

- 3.2 For Q fever Between 10 and 30 days after the final embryo collection, the donor females were tested with negative results for Q fever using the complement fixation test (CFT) (negative being no fixation of complement at a dilution of 1:10 or higher) or the ELISA

Test used:

Date(s) of sample collection:

- 3.3 All testing was conducted at a laboratory approved by the Department to conduct export testing, and laboratory results for tests specified in this certificate are attached.

4 Storage and transport

- 4.1 All straws are clearly marked with the identification of the donor animals and the date(s) of collection. If a code is used for this information, its decipher must accompany the consignment.

- 4.2 The embryos were only stored with other embryos or semen that were eligible for export to New Zealand. The containers were held in an approved storage place under the supervision of the Department until export.

- 4.3 The embryos were placed in new or sterilised transport containers filled with fresh (previously unused) liquid nitrogen. Tank serial N^o:

Method of sterilisation (if applicable):

Date of sterilisation (if applicable):

- ~~4.4 Prior to export, the container in which the embryos are to be transported was sealed by either the embryo collection team veterinarian or an Official Veterinarian using seals bearing the marks:~~

Schedule 1:**I. Information Concerning the Donor Animals (Females and Males)**

Donor Animals				
	Breed	Identification	Date of birth	Batch number of semen (if applicable) or date of natural mating
Donor Ewes/Doe				
Donor Rams/buck				

II. Information Concerning the Ovine/Caprine Embryos

Collection Date	Female ID	Male ID	Straw Identification (markings to be indelible)	Nº Straws per donor (include Nº Embryos per straw)

SCHEDULE OF TESTING

Ewes/Doe	Identification	Q Fever Test Type	Date Of Test	Result

Ewes/Doe	Identification	Bluetongue Test Type	Date Of Test	Result

Signature (pdf. doc only)

Date